

## Review Article

# RNA-Binding Proteins in Amyotrophic Lateral Sclerosis and Neurodegeneration

Scott E. Ugras<sup>1,2</sup> and James Shorter<sup>1,2</sup>

<sup>1</sup>Biochemistry and Molecular Biophysics Graduate Group, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA 19104, USA

<sup>2</sup>Department of Biochemistry and Biophysics, Perelman School of Medicine, University of Pennsylvania, 805b Stellar-Chance Laboratories, 422 Curie Boulevard, Philadelphia, PA 19104, USA

Correspondence should be addressed to James Shorter, jshorter@mail.med.upenn.edu

Received 26 January 2012; Accepted 26 February 2012

Academic Editor: Jeff Bronstein

Copyright © 2012 S. E. Ugras and J. Shorter. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Amyotrophic Lateral Sclerosis (ALS) is an adult onset neurodegenerative disease, which is universally fatal. While the causes of this devastating disease are poorly understood, recent advances have implicated RNA-binding proteins (RBPs) that contain predicted prion domains as a major culprit. Specifically, mutations in the RBPs TDP-43 and FUS can cause ALS. Cytoplasmic mislocalization and inclusion formation are common pathological features of TDP-43 and FUS proteinopathies. Though these RBPs share striking pathological and structural similarities, considerable evidence suggests that the ALS-linked mutations in TDP-43 and FUS can cause disease by disparate mechanisms. In a recent study, Couthouis et al. screened for protein candidates that were also involved in RNA processing, contained a predicted prion domain, shared other phenotypic similarities with TDP-43 and FUS, and identified TAF15 as a putative ALS gene. Subsequent sequencing of ALS patients successfully identified ALS-linked mutations in TAF15 that were largely absent in control populations. This study underscores the important role that perturbations in RNA metabolism might play in neurodegeneration, and it raises the possibility that future studies will identify other RBPs with critical roles in neurodegenerative disease.

## 1. Introduction

Amyotrophic Lateral Sclerosis (ALS) is an adult-onset neurodegenerative disease that is typically fatal within 2–5 years of disease onset. Loss of upper and lower motor neurons leads to paralysis and is usually the cause of death. Most cases are sporadic with no family history, but ~10% of cases are familial [1].

Efforts to understand the causes of ALS and methods to mitigate its pathogenesis have been severely limited by an inability to identify ALS genes. For the past two decades, most ALS research has focused on the ubiquitously expressed copper/zinc superoxide dismutase 1 (SOD1). Mutations in *SOD1* underlie ~20% of familial ALS (fALS) cases and ~1% of sporadic ALS (sALS) cases [1]. No consensus has emerged on the mechanism by which these SOD1 mutations lead to selective neuronal death in ALS. However, loss of SOD1

function does not cause motor dysfunction and ALS-linked SOD1 mutants can have wild-type enzyme activity [2]. Thus, a toxic gain of function of misfolded SOD1 likely underpins pathogenesis [2]. Recently, a number of genes involved in RNA processing have been identified that can cause ALS in select cases [3–7]. These findings have revolutionized our understanding of ALS and have simultaneously changed the genetic landscape of neurodegenerative disease.

## 2. TDP-43 Is an ALS Disease Protein

The identification of a novel ALS disease protein in 2006 dramatically altered our understanding of this disease and brought to light the potential role RNA-binding proteins (RBPs) play in neurodegeneration. TDP-43 was found in cytoplasmic inclusions in diseased brain [3]. Shortly thereafter, many mutations in *TDP43* were found in sALS and

fALS cases, but not in control cases [6, 7]. This finding took the ALS community by surprise and caused many researchers to focus on TDP-43 and the role other RBPs might play in ALS.

TDP-43 is a highly conserved, ubiquitously expressed protein that has various roles in RNA metabolism [8, 9]. TDP-43 contains two RNA Recognition Motifs (RRMs) and a C-terminal Glycine-rich domain [1, 8, 10]. While TDP-43 is predominantly a nuclear protein, in ALS motor neurons it was found to mislocalize to the cytoplasm, with some affected neurons even showing complete clearance of TDP-43 from the nucleus [3]. In addition to this mislocalization, TDP-43 was also found to be modified and to be misfolded in an aggregated state [3]. Its modifications include ubiquitination, phosphorylation, and cleavage [3, 11]. The implications of these modifications as well as their origins and roles in ALS pathology, remain largely unclear and require further investigation [11].

### 3. FUS Is Also an ALS Disease Protein

The identification of the RBP, TDP-43, as a causative agent of ALS-directed research towards the possibility that other RBPs might also play a role in ALS. This led to the contemporaneous discovery by two groups of the ALS disease protein Fused in Sarcoma, or FUS [4, 5]. Like TDP-43, FUS is a ubiquitously expressed protein that is normally nuclear but is found in cytoplasmic inclusions in ALS brain [4, 5].

FUS contains an N-terminal QGSY-rich domain, followed by a Glycine-rich domain, an RRM, and two RGG-rich domains [1, 10]. Like TDP-43, FUS has roles in a number of different RNA-processing activities, including splicing and mRNA export to the cytoplasm [10]. To date, about 50 ALS-linked FUS mutations have been identified [10].

### 4. TDP-43 and FUS Are Highly Prone to Aggregation and Contain Predicted Prion Domains

Prions are infectious proteins that are prone to misfold and aggregate and can spread from organism to organism in a DNA/RNA-independent manner [12–17]. They accomplish this by converting natively folded conformers with the same primary sequence into the prion state [14, 15]. Prions cause a number of deadly mammalian diseases, such as bovine spongiform encephalopathy and Creutzfeldt-Jakob disease in humans [12]. A similar process to the spreading of prions might be involved in the cell-to-cell transmission of disease proteins in a number of neurodegenerative diseases, including Alzheimer's disease, Parkinson's disease, and Huntington's disease [13, 16–18].

TDP-43 and FUS both contain a predicted prion domain similar to those found in yeast prion proteins like Sup35 and Ure2 [13, 16, 18–20]. Prion domains confer prion behavior by transitioning from an intrinsically unfolded conformation to a variety of infectious amyloid forms [13–16, 18]. In TDP-43 the predicted prion domain (amino acids 277–414) largely overlaps with the C-terminal Glycine-rich domain,

whereas in FUS it resides in the N-terminal QGSY-rich and Glycine-rich domains (amino acids 1–239) [13, 16, 18]. The presence of predicted prion domains and the incorporation of these RBPs into disease-associated inclusions suggested that these RBPs are highly prone to aggregation [13, 16, 18]. It should be noted, however, that neither TDP-43 nor FUS aggregates have been shown to harbor infectious properties, that is, to be self-templating conformers that are naturally transmitted between individuals and promote a self-perpetuating phenotypic change [16]. Indeed, it is not yet clear whether full-length FUS or TDP-43 aggregates are self-templating [16, 21, 22], although specific TDP-43 fragments that harbor the predicted prion domain can access self-templating forms [23]. However, an interesting aspect of ALS is the spreading of pathology to neighboring anatomic regions, particularly in the brain [24]. While this suggests that prion-like behavior is involved in ALS, a more direct demonstration that TDP-43 and FUS can act as bona fide prions is currently lacking.

To test the prediction that these RBPs are intrinsically aggregation-prone, a cell-free system was established to monitor the propensity of pure protein for spontaneous aggregation [21, 22, 25]. Both TDP-43 and FUS were found to rapidly aggregate in vitro, with a lag phase that was either on the order of minutes or too short to measure [21, 22, 25]. Interestingly, both proteins form pore-shaped oligomers and filamentous aggregates that closely resemble the structure of pathological aggregates detected in motor neurons of ALS patients [21, 22, 25].

### 5. Yeast Can Be a Useful System to Model Protein Misfolding Connected with Neurodegeneration

The baker's yeast *Saccharomyces cerevisiae* has been extensively used to gain insight into many neurodegenerative processes [26]. Furthermore, since disease proteins recapitulate many aspects of pathogenesis in yeast, studying TDP-43 and FUS in yeast was an immediately attractive option [26]. Importantly, overexpression of either TDP-43 or FUS in yeast results in two key aspects also seen in human cases: cytoplasmic aggregation and toxicity [21, 22, 27–34]. This finding suggests that yeast can be used as a model system to study the misfolding and toxicity of ALS proteins.

### 6. ALS-Linked Mutations in TDP-43 and FUS Are Likely to Cause Disease by Distinct Mechanisms

Though TDP-43 and FUS share many striking similarities [10], several lines of evidence indicate that their respective ALS-linked mutations cause disease by distinct pathways [18]. Many of the TDP-43 ALS-linked mutations localize to the C-terminal Glycine-rich domain [10], which is thought to be critical for TDP-43 misfolding since this is also the location of the predicted prion domain [13, 21, 29]. On the other hand, most of the FUS ALS-linked mutations reside in the extreme C-terminal region, which is also the location of

the nuclear localization signal (NLS) of FUS [10]. FUS has a nonclassical NLS comprised of highly conserved proline and tyrosine residues (PY-NLS), which is decoded by the nuclear import factor Transportin-1 (TNPO1) [35].

Most of the TDP-43 ALS-linked mutations tested to date accelerate aggregation *in vitro*, whereas the FUS mutations have no effect [21, 22]. Q331K and M337V, for instance, are particularly aggressive TDP-43 variants that aggregate more rapidly than WT [21]. The FUS mutants H517Q, R521C, and R521H do not affect the aggregation kinetics of FUS [22]. The presence of the FUS mutations in the PY-NLS suggested that these could disrupt its cellular localization rather than its aggregation [35, 36]. To test this prediction, GFP-tagged WT-FUS as well as C-terminal FUS mutants were expressed in a mammalian cell line and FUS localization was examined [36]. The C-terminal mutations, including R521C, R524S, R522G, and P525L, caused FUS mislocalization to the cytoplasm, in a manner dependent on the severity of the mutations in altering the integrity of the PY-NLS [35, 36]. Interestingly, the degree of cytoplasmic mislocalization was correlated with mean age of disease onset, with mutations (e.g., P525L) causing the greatest cytoplasmic mislocalization also resulting in the earliest age of disease onset [36]. Furthermore, knockdown or inhibition of TNPO1 also resulted in WT-FUS cytoplasmic mislocalization, indicating that the FUS-TNPO1 interaction is crucial for proper cellular localization [36]. Together, these data provide strong evidence that specific ALS-linked TDP-43 mutations primarily cause disease by promoting aggregation, whereas specific ALS-linked FUS mutations cause disease by perturbing nuclear localization [18].

## 7. A Yeast Screen Identified TAF15 as a Putative ALS Gene

Given the largely overlapping functions and protein domains of TDP-43 and FUS, it is likely that many other RBPs play a crucial role not only in ALS but also in neurodegeneration more generally [16, 18, 25]. This tantalizing possibility led Couthouis et al. to perform a yeast functional screen in an attempt to harness the power of yeast to identify new candidate ALS disease genes [25].

First, the entire human genome was surveyed for proteins that contained RRM [25]. Of the 213 human genes containing RRM, Couthouis et al. were able to clone 133 of them into yeast expression vectors as YFP fusion proteins [25]. Next, to narrow the list to those proteins that had similar phenotypes to TDP-43 and FUS in yeast, 38 were identified that were toxic and formed cytoplasmic foci [25]. Of these 38, a bioinformatics approach was taken to rank these in order of strongest to weakest predicted prion domains [25]. 13 of these 38 proteins contain a predicted prion domain and at the top of this list sat TAF15 [25].

Interestingly, TAF15 shares many similarities with TDP-43 and FUS [16]. Its domain architecture is very similar to FUS; it contains an N-terminal QGSY-rich domain, a Glycine-rich domain, an RRM, two RGG domains, and a C-terminal PY-NLS. Like FUS, it contains a predicted prion

domain (amino acids 1–152) that overlaps with its QGSY-rich domain [16]. Furthermore, pure TAF15 also aggregates with extremely rapid kinetics and forms aggregates that are morphologically similar to those formed by FUS [25]. Sequencing of the TAF15 gene in ALS and control populations led to the successful identification of several mutations enriched in ALS patients, including M368T, G391E, R408C, G452E, and G473E, which are absent in control populations [25]. Furthermore, most of these mutations cause TAF15 to aggregate more rapidly *in vitro* and *in vivo*, similar to TDP-43 ALS-linked mutations [25]. Although these specific TAF15 variants have not been connected to fALS, the search is ongoing [25]. Nonetheless, this study underscores the power of yeast as a model system to study the misfolding of disease-associated proteins and the potentially general role of RBPs in neurodegeneration [25].

## 8. Conclusions

The identification of TDP-43 as an ALS gene altered the genetic landscape of neurodegenerative disease. The importance of altered RNA processing in disease pathogenesis is now much more widely appreciated. Furthermore, the identification of other RBPs that all contain RRM and prion domains is unlikely to be mere coincidence [16, 18]. Indeed, it is highly likely that future research will identify many other RBPs and related proteins in neurodegeneration [16]. For instance, recent evidence has linked EWSR1, another RBP very similar to FUS and TAF15, to frontotemporal lobar degeneration with ubiquitin-positive inclusions (FTLD-U) [37]. Additionally, it is likely that the DNA or RNA targets of these RBPs play a significant role in disease [33, 38–41]. Remarkably, a hexanucleotide intronic repeat was recently identified that can account for a large fraction of ALS cases [42, 43]. Further examination and elucidation of these processes involving RBPs in disease will be critical.

Though much progress has been made in the identification and characterization of the roles that RBPs play in ALS, much is still beyond our current understanding. For instance, it is still not known whether these proteins cause disease by a loss of function or gain of function or both [1, 10, 11, 16–18]. It is still unclear how certain ALS-linked mutations cause disease. For example, some TDP-43 ALS-linked mutations do not promote aggregation [21] and several FUS ALS-linked mutations lie outside of the PY-NLS [10]. It is unclear whether these mutations cause FUS to aggregate more rapidly, to interact with other proteins, or to disrupt its normal RNA targets. The role of specific RNA targets with which these RBPs interact may also be key to disease pathogenesis [38–41]. Ultimately, an enhanced understanding of the exact pathways these RBPs take en route to disease onset is required to truly understand pathogenesis and to develop effective therapeutic remedies.

## Acknowledgments

The authors would like to thank Dr. Janis Burkhardt and Dr. Aaron Gitler for reading over this paper and for providing critical feedback. J. Shorter is supported by an NIH

Director's New Innovator Award (1DP2OD002177-01), an Ellison Medical Foundation New Scholar in Aging Award, and by The Robert Packard Center for ALS Research at Johns Hopkins.

## References

- [1] C. Lagier-Tourenne and D. W. Cleveland, "Rethinking ALS: the FUS about TDP-43," *Cell*, vol. 136, no. 6, pp. 1001–1004, 2009.
- [2] H. Ilieva, M. Polymenidou, and D. W. Cleveland, "Non-cell autonomous toxicity in neurodegenerative disorders: ALS and beyond," *Journal of Cell Biology*, vol. 187, no. 6, pp. 761–772, 2009.
- [3] M. Neumann, D. M. Sampathu, L. K. Kwong et al., "Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis," *Science*, vol. 314, no. 5796, pp. 130–133, 2006.
- [4] T. J. Kwiatkowski Jr., D. A. Bosco, A. L. LeClerc et al., "Mutations in the FUS/TLS gene on chromosome 16 cause familial amyotrophic lateral sclerosis," *Science*, vol. 323, no. 5918, pp. 1205–1208, 2009.
- [5] C. Vance, B. Rogelj, T. Hortobagyi et al., "Mutations in FUS, an RNA processing protein, cause familial amyotrophic lateral sclerosis type 6," *Science*, vol. 323, no. 5918, pp. 1208–1211, 2009.
- [6] E. Kabashi, P. N. Valdmanis, P. Dion et al., "TARDBP mutations in individuals with sporadic and familial amyotrophic lateral sclerosis," *Nature Genetics*, vol. 40, no. 5, pp. 572–574, 2008.
- [7] J. Sreedharan, I. P. Blair, V. B. Tripathi et al., "TDP-43 mutations in familial and sporadic amyotrophic lateral sclerosis," *Science*, vol. 319, no. 5870, pp. 1668–1672, 2008.
- [8] E. Buratti and F. E. Baralle, "The multiple roles of TDP-43 in pre-mRNA processing and gene expression regulation," *RNA Biology*, vol. 7, no. 4, pp. 420–429, 2010.
- [9] Y. Kawahara and A. Mieda-Sato, "TDP-43 promotes microRNA biogenesis as a component of the Drosha and Dicer complexes," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 109, no. 9, pp. 3347–3352, 2012.
- [10] S. da Cruz and D. W. Cleveland, "Understanding the role of TDP-43 and FUS/TLS in ALS and beyond," *Current Opinion in Neurobiology*, vol. 21, no. 6, pp. 904–919, 2011.
- [11] E. B. Lee, V. M. Lee, and J. Q. Trojanowski, "Gains or losses: molecular mechanisms of TDP43-mediated neurodegeneration," *Nature Reviews Neuroscience*, vol. 13, no. 1, pp. 38–50, 2012.
- [12] D. W. Colby and S. B. Prusiner, "De novo generation of prion strains," *Nature Reviews Microbiology*, vol. 9, no. 11, pp. 771–777, 2011.
- [13] M. Cushman, B. S. Johnson, O. D. King, A. D. Gitler, and J. Shorter, "Prion-like disorders: blurring the divide between transmissibility and infectivity," *Journal of Cell Science*, vol. 123, no. 8, pp. 1191–1201, 2010.
- [14] J. Shorter, "Emergence and natural selection of drug-resistant prions," *Molecular Biosystems*, vol. 6, no. 7, pp. 1115–1130, 2010.
- [15] J. Shorter and S. Lindquist, "Prions as adaptive conduits of memory and inheritance," *Nature Reviews Genetics*, vol. 6, no. 6, pp. 435–450, 2005.
- [16] O. D. King, A. D. Gitler, and J. Shorter, "The tip of the iceberg: RNA-binding proteins with prion-like domains in neurodegenerative disease," *Brain Research*. In press.
- [17] M. Polymenidou and D. W. Cleveland, "The seeds of neurodegeneration: prion-like spreading in ALS," *Cell*, vol. 147, no. 3, pp. 498–508, 2011.
- [18] A. D. Gitler and J. Shorter, "RNA-binding proteins with prion-like domains in ALS and FTL-D-U," *Prion*, vol. 5, no. 3, pp. 179–187, 2011.
- [19] R. A. Fuentealba, M. Udan, S. Bell et al., "Interaction with polyglutamine aggregates reveals a Q/N-rich domain in TDP-43," *Journal of Biological Chemistry*, vol. 285, no. 34, pp. 26304–26314, 2010.
- [20] M. Udan and R. H. Baloh, "Implications of the prion-related Q/N domains in TDP-43 and FUS," *Prion*, vol. 5, no. 1, pp. 1–5, 2011.
- [21] B. S. Johnson, D. Snead, J. J. Lee, J. M. McCaffery, J. Shorter, and A. D. Gitler, "TDP-43 is intrinsically aggregation-prone, and amyotrophic lateral sclerosis-linked mutations accelerate aggregation and increase toxicity," *Journal of Biological Chemistry*, vol. 284, no. 30, pp. 20329–20339, 2009.
- [22] Z. Sun, Z. Diaz, X. Fang et al., "Molecular determinants and genetic modifiers of aggregation and toxicity for the ALS disease protein FUS/TLS," *PLoS Biology*, vol. 9, no. 4, Article ID e1000614, 2011.
- [23] Y. Furukawa, K. Kaneko, S. Watanabe, K. Yamanaka, and N. Nukina, "A seeding reaction recapitulates intracellular formation of sarkosyl-insoluble transactivation response element (TAR) DNA-binding protein-43 inclusions," *Journal of Biological Chemistry*, vol. 286, no. 21, pp. 18664–18672, 2011.
- [24] J. M. Ravits and A. R. La Spada, "ALS motor phenotype heterogeneity, focality, and spread: deconstructing motor neuron degeneration," *Neurology*, vol. 73, no. 10, pp. 805–811, 2009.
- [25] J. Couthouis, M. P. Hart, J. Shorter et al., "A yeast functional screen predicts new candidate ALS disease genes," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 108, no. 52, pp. 20881–20890, 2011.
- [26] A. D. Gitler, "Beer and bread to brains and beyond: can yeast cells teach us about neurodegenerative disease?" *Neurosignals*, vol. 16, no. 1, pp. 52–62, 2008.
- [27] M. Armakola, M. P. Hart, and A. D. Gitler, "TDP-43 toxicity in yeast," *Methods*, vol. 53, no. 3, pp. 238–245, 2011.
- [28] R. J. Braun, C. Sommer, D. Carmona-Gutierrez et al., "Neurotoxic 43-kDa TAR DNA-binding protein (TDP-43) triggers mitochondrion-dependent programmed cell death in yeast," *Journal of Biological Chemistry*, vol. 286, no. 22, pp. 19958–19972, 2011.
- [29] B. S. Johnson, J. M. McCaffery, S. Lindquist, and A. D. Gitler, "A yeast TDP-43 proteinopathy model: exploring the molecular determinants of TDP-43 aggregation and cellular toxicity," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 105, no. 17, pp. 6439–6444, 2008.
- [30] D. Kryndushkin, R. B. Wickner, and F. Shewmaker, "FUS/TLS forms cytoplasmic aggregates, inhibits cell growth and interacts with TDP-43 in a yeast model of amyotrophic lateral sclerosis," *Protein and Cell*, vol. 2, no. 3, pp. 223–236, 2011.
- [31] S. Ju, D. F. Tardiff, H. Han et al., "A yeast model of FUS/TLS-dependent cytotoxicity," *PLoS Biology*, vol. 9, no. 4, Article ID e1001052, 2011.
- [32] K. Fushimi, C. Long, N. Jayaram, X. Chen, L. Li, and J. Y. Wu, "Expression of human FUS/TLS in yeast leads to protein

- aggregation and cytotoxicity, recapitulating key features of FUS proteinopathy,” *Protein and Cell*, vol. 2, no. 2, pp. 141–149, 2011.
- [33] A. C. Elden, H. J. Kim, M. P. Hart et al., “Ataxin-2 intermediate-length polyglutamine expansions are associated with increased risk for ALS,” *Nature*, vol. 466, no. 7310, pp. 1069–1075, 2010.
- [34] D. F. Tardiff, M. L. Tucci, K. A. Caldwell, G. A. Caldwell, and S. Lindquist, “Different 8-hydroxyquinolines protect models of TDP-43 protein, alpha-synuclein, and polyglutamine proteotoxicity through distinct mechanisms,” *Journal of Biological Chemistry*, vol. 287, no. 6, pp. 4107–4120, 2012.
- [35] B. J. Lee, A. E. Cansizoglu, K. E. Suel, T. H. Louis, Z. Zhang, and Y. M. Chook, “Rules for nuclear localization sequence recognition by karyopherin  $\beta 2$ ,” *Cell*, vol. 126, no. 3, pp. 543–558, 2006.
- [36] D. Dormann, R. Rodde, D. Edbauer et al., “ALS-associated fused in sarcoma (FUS) mutations disrupt transportin-mediated nuclear import,” *EMBO Journal*, vol. 29, no. 16, pp. 2841–2857, 2010.
- [37] M. Neumann, E. Bentmann, D. Dormann et al., “FET proteins TAF15 and EWS are selective markers that distinguish FTLD with FUS pathology from amyotrophic lateral sclerosis with FUS mutations,” *Brain*, vol. 134, no. Pt9, pp. 2595–2609, 2011.
- [38] J. I. Hoell, E. L. Larsson, S. Runge et al., “RNA targets of wild-type and mutant FET family proteins,” *Nature Structural and Molecular Biology*, vol. 18, no. 12, pp. 1428–1431, 2011.
- [39] M. Polymenidou, C. Lagier-Tourenne, K. R. Hutt et al., “Long pre-mRNA depletion and RNA missplicing contribute to neuronal vulnerability from loss of TDP-43,” *Nature Neuroscience*, vol. 14, no. 4, pp. 459–468, 2011.
- [40] Y. M. Ayala, L. de Conti, S. E. Avendano-Vazquez et al., “TDP-43 regulates its mRNA levels through a negative feedback loop,” *The EMBO Journal*, vol. 30, no. 2, pp. 277–288, 2011.
- [41] J. R. Tollervy, T. Curk, B. Rogelj et al., “Characterizing the RNA targets and position-dependent splicing regulation by TDP-43,” *Nature Neuroscience*, vol. 14, no. 4, pp. 452–458, 2011.
- [42] M. DeJesus-Hernandez, I. R. Mackenzie, B. F. Boeve et al., “Expanded GGGGCC hexanucleotide repeat in noncoding region of C9ORF72 causes chromosome 9p-linked FTD and ALS,” *Neuron*, vol. 72, no. 2, pp. 245–256, 2011.
- [43] A. E. Renton, E. Majounie, A. Waite et al., “A hexanucleotide repeat expansion in C9ORF72 is the cause of chromosome 9p21-linked ALS-FTD,” *Neuron*, vol. 72, no. 2, pp. 257–268, 2011.